



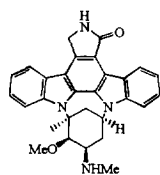
THE PREPARATION AND SAR OF 4-(ANILINO), 4-(PHENOXY), AND 4-(THIOPHENOXY)-QUINAZOLINES: INHIBITORS OF p56^{lck} AND EGF-R TYROSINE KINASE ACTIVITY.

Michael R. Myers,^{a*} Natalie N. Setzer,^a Alfred. P. Spada,^a Allison L. Zulli,^a Chin-Yi J. Hsu,^b Asher Zilberstein,^b Susan E. Johnson,^b Linda E. Hook,^b and Mary V. Jacoski.^b

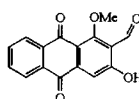
*Departments of Medicinal Chemistry^a and Inflammation & Bone Metabolism^b
Rhône-Poulenc Rorer Pharmaceuticals, 500 Arcola Rd. Collegeville, PA 19426-0107 U.S.A.*

Abstract: We report herein our preliminary results of a SAR study of quinazoline-based inhibitors of p56^{lck} and EGF-R tyrosine kinase activity.¹ The most potent inhibitor of p56^{lck} identified, RPR-108518A (10), has an IC₅₀ of 0.50 μM. The 3-chlorophenoxy- and 3-chlorothiophenoxy- derivatives 5 and 6 were also shown to be extremely potent EGF-R inhibitors. © 1997, Elsevier Science Ltd. All rights reserved.

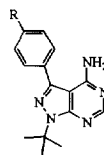
Inhibitors of p56^{lck} tyrosine kinase² reported in the literature include compounds that are potent but nonselective (staurosporine,³ competitive with ATP) or compounds that are very weak tyrosine kinase inhibitors but are somewhat selective (flavonoids, i.e., quercetin⁴). In addition, NCI workers disclosed a new series of dihydroxyisoquinolines⁵ that have potent p56^{lck} inhibitory activity. Damnacanthal⁶ was also reported to be a selective and potent (17 nM) inhibitor of p56^{lck}. A group at Pfizer recently disclosed two novel potent inhibitors⁷ of p56^{lck}; PP1 and PP2 have IC₅₀s of 5 nM and 4 nM, respectively. Potential therapeutic uses for selective inhibitors of p56^{lck} include the treatment of autoimmune diseases such as rheumatoid arthritis or transplant rejection.



Staurosporine

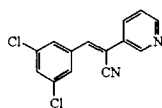


Damnacanthal

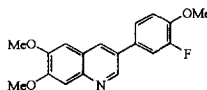


PP1, R = CH₃
PP2, R = Cl

Workers at Zeneca⁸ and Parke-Davis⁹ have published a number of reports describing exceptionally potent quinazoline-based epidermal growth factor receptor (EGF-R) inhibitors. Recently a Ciba-Geigy group disclosed a series of bioisosteric pyrrolo-pyrimidines that demonstrate potent EGF-R activity.¹⁰ Selective inhibition of p56^{lck} tyrosine kinase has not been reported for the quinazoline class of inhibitors. Our group had previously identified a series of potent and selective inhibitors for both EGF-R^{11a} and platelet-derived growth factor receptor (PDGF-R) tyrosine kinases (1 and 2, respectively).^{11b,c} These compounds and others from our corporate database were evaluated in an assay for p56^{lck} inhibition. This report describes our preliminary efforts toward optimizing quinazoline-based leads that we identified early in our screening program.

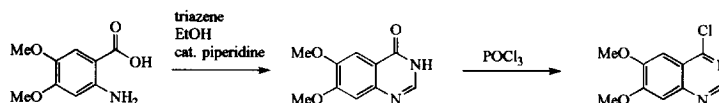


1



2

The 4-chloroquinazoline starting materials were prepared from the commercially available anthranilic acids via treatment with formamide in the usual way¹² followed by reaction with POCl₃ at reflux for 3 h. Aqueous workup followed by flash chromatography provided the requisite 4-chloroquinazolines. We found that by using the closure conditions of Kreutzberger and Uzbek¹³ (triazene, catalytic piperidine) we were able to prepare pure 6,7-dimethoxyquinazolinone in very high yield. This route is preferred over the classical formamide route since the quinazolinone is cleanly converted to the 4-chloroquinazoline with POCl₃ on large scale with the avoidance of column chromatography.



The targeted anilinoquinazolines were prepared by simply heating the appropriate chloroquinazoline with an aniline derivative in ethanol for 5-15 min. The resulting precipitate was isolated directly as the HCl salt and was dried in vacuo to provide analytically pure material. Alternatively, the phenoxy- and thiophenoxy- derivatives were prepared via NaH treatment of the corresponding phenol or thiophenol in THF followed by addition of the chloroquinazoline, workup and column chromatography.

We have confirmed that **3**, **4** and **11** are potent inhibitors of EGF-R tyrosine kinase activity as reported.^{8,9} As can be seen in Table 1, **3** does not demonstrate significant activity in our p56^{lck} assay.¹⁴ However, addition of the 6,7-dimethoxy groups provided compound **4**, which is a moderately active inhibitor of p56^{lck} tyrosine kinase activity. Replacement of the nitrogen-linker in the *m*-chlorophenyl- derivative **4** with oxygen (**5**), or sulfur (**6**), led to a small improvement in p56^{lck} and EGF-R activity. Optimal inhibition of p56^{lck} tyrosine kinase activity was obtained with the 3,4,5-trimethoxyanilino derivative **10**. Removal of one or two methoxy substituents at any of these positions reduced activity (**9** or **16** or **17** vs **10**). In the case of the dimethoxy-derivative **17**, replacement of the amine-linker with sulfur (**18**) resulted in a loss of activity for both p56^{lck} and EGF-R, contrasting the SAR seen with the meta-chlorophenyl series **4-6**. Attempts to prepare compounds with alternative electron-donating substituents did not lead to any improvement in activity or selectivity vs. EGF-R (**14**, **15**). Substitution of the aniline with a *N*-methyl aniline eliminated p56^{lck} inhibitory activity (**7** vs **8**).

In an effort to identify other features of the 6,7-dimethoxyquinazoline that are important to the SAR for p56^{lck} we prepared a series of substituted quinazolines (**19-23**) while keeping the 3,4,5-trimethoxyanilino-substituent constant. Removal of a single methoxy group (**22**) results in a 20-fold loss in activity. Substitution of the amine-linker with oxygen (**10** vs. **24**) led to a 10-fold loss in activity, which contradicts the slight improvement in activity seen with the *m*-chlorophenyl- derivatives **4** and **5**. The allopurinol and adenine derivatives did not afford any significant improvement in activity (**25**, **26**).

Particularly striking is the lack of activity observed with compounds substituted at the 2- and 8-positions of the quinazoline (**19-21**). This SAR parallels our own experience with the quinoline-based inhibitors of PDGF-R^{11c} and quinazoline-based inhibitors of CSF-1R¹⁵ and highlights the importance of steric hinderance in the *N*-1 interaction with the enzyme. A binding model using bioisosteric pyrrolo-pyrimidine EGF-R inhibitors has been proposed¹⁰ that is consistent with the dramatic loss of activity seen with **19-21**. In the case of PDGF-R tyrosine kinase inhibitors like **2**, it was found that a similarly-substituted naphthalene was not an acceptable substitute for the quinoline moiety.^{11c} Burke has observed similar results with isoquinoline inhibitors of p56^{lck}.⁵

In regards to EGF-R activity, compounds **5** and **6** were found to be exceptionally potent inhibitors. Comparison of the *m*-chlorophenylthio- group in **6** with the 3,4-dimethoxyphenylthio- group in **18** shows complete loss of EGF-R activity. However, there is only a 10-fold loss between the similarly substituted amine-linked derivatives

4 and **17**. Interestingly, compound **8** has an IC_{50} of only 4.0 μM for inhibition of EGF-R autophosphorylation vs **7**, which has an IC_{50} of 0.05 μM . This loss of potency is likely due in part to the conformation of the phenyl group in relation to the quinazoline ring¹⁵ and not to the loss of a specific NH interaction since the oxygen and sulfur analogues (**5** and **6**) retain potent activity against EGF-R.

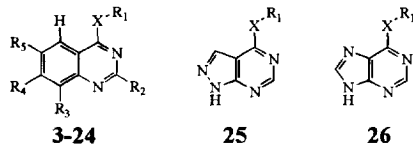


Table 1.

Cmpd	X	R1	R2	R3	R4	R5	p56 ^{lck} IC ₅₀ (μM)	EGF-R IC ₅₀ (μM)	mp	CHN ^b
3	NH	3-Cl-phenyl	H	H	H	H	50	0.05-0.10	245-50 ^a	HCl
4	NH	3-Cl-phenyl	H	H	OMe	OMe	5	0.03	261-65 ^a	HCl
5	O	3-Cl-phenyl	H	H	OMe	OMe	1	0.02	152-53	---
6	S	3-Cl-phenyl	H	H	OMe	OMe	2.5	0.01	152-53.5	---
7	NH	phenyl	H	H	OMe	OMe	10	0.05	264-66 ^a	HCl
8	NMe	phenyl	H	H	OMe	OMe	>100	4.0	233-37 ^a	HCl, 0.8 M EtOH
9	NH	3,5-diMeO-phenyl	H	H	OMe	OMe	10	3.0	270-75 ^a	HCl
10	NH	3,4,5-triMeO-phenyl	H	H	OMe	OMe	0.50	0.50	260-65 ^a	HCl
11	NH	benzyl	H	H	OMe	OMe	20	0.004	220-25 ^a	HCl, 0.2 M H ₂ O
12	NH	3-F-phenyl	H	H	OMe	OMe	10	0.025	270-72 ^a	HCl
13	NH	5-indanyl	H	H	OMe	OMe	50	0.60	244-46 ^a	HCl, 1.4 M H ₂ O
14	NH	4-hydroxyphenyl	H	H	OMe	OMe	3	0.10	253-58 ^a	HCl, 0.9 M H ₂ O
15	NH	4-N-morpholinophenyl	H	H	OMe	OMe	>100	>1 ^c	231-35 ^a	HCl, 1.0 M H ₂ O
16	NH	4-MeO-phenyl	H	H	OMe	OMe	10	0.25	220-30 ^a	HCl
17	NH	3,4-diMeO-phenyl	H	H	OMe	OMe	10	0.35	272-75 ^a	HCl
18	S	3,4-diMeO-phenyl	H	H	OMe	OMe	>50	>50	196-99	---
19	NH	3,4,5-triMeO-phenyl	H	Me	H	Me	>100	>20	275-78 ^a	HCl
20	NH	3,4,5-triMeO-phenyl	H	OMe	OMe	OMe	>100	>20	235 ^a	HCl
21	NH	3,4,5-triMeO-phenyl	Cl	H	OMe	OMe	>100	>20	126-30	HCl, 1.0 M H ₂ O
22	NH	3,4,5-triMeO-phenyl	H	H	H	OMe	10	0.10	235-37 ^a	HCl, 1.0 M H ₂ O
23	NH	3,4,5-triMeO-phenyl	H	H	H	Cl	>50	>1 ^c	257-61 ^a	HCl
24	O	3,4,5-triMeO-phenyl	H	H	OMe	OMe	5	NT ^d	228-32	---
25	NH	3,4,5-triMeO-phenyl	1H-pyrazolo[3,4-d]pyrimidin-4-yl				10	NT ^d	250-52 ^a	HCl
26	NH	3,4,5-triMeO-phenyl	9H-purinyl-6-yl				50	NT ^d	>250	HCl

(a) decomposed. (b) CHN experimentally determined (with indicated amount of associated solvent or water) to be within ± 0.3 of the theoretical value. (c) Single experiment, did not reach IC_{50} value. (d) Not tested

In summary, we have found that RPR-108518A (10) is a moderate nonselective inhibitor of p56^{lck} tyrosine kinase. The accompanying paper demonstrates that fine-tuning for selectivity vs other tyrosine kinases is possible using a quinazoline as a template. Further work towards identifying compounds with improved in vitro activity and selectivity will be reported separately. Preliminary results using RPR-108518A in intact-cell studies¹⁶ show that inhibition of p56^{lck} tyrosine kinase activity diminishes the levels of substrate tyrosine phosphorylation upon stimulation; IL-2 secretion is also inhibited in Jurkat cells with an IC₅₀ that corresponds to the inhibition of in situ p56^{lck} autophosphorylation.

Acknowledgment: The authors wish to acknowledge Paul Persons for the preparation of compound 18.

References:

1. Partially disclosed in Myers, M. R.; Spada, A. P.; Maguire, M. P.; Persons, P. E.; Zilberstein, A.; Hsu, C.-Y.; Johnson, S., WO PCT Appl. 95/15758, 1995.
2. Bolen, J. B.; Rowley, R. B.; Spana, C.; Tsygankov, A. Y. *FASEB J.* **1992**, 3403. Mustelin, T.; Burn, P. *TIBS* **1993**, 215. Eichmann, K. *Angew. Chem. Int. Ed. Eng.* **1993**, 54. Klausner, R. D.; Samelson, L. E. *Cell* **1991**, 875.
3. Tamaoki, T.; Nomoto, H.; Takahashi, I.; Kato, Y.; Morimoto, M.; Tomita, F. *Biochem. Biophys. Res. Comm.* **1986**, 135, 397. Herbert, J. M.; Seban, E.; Maffrand, J. P. *Biochem. Biophys. Res. Comm.* **1990**, 171, 189.
4. Hagiwara, M.; Inoue, S.; Tamaka, T.; Nunoki, K.; Ito, M.; Hidaka, H. *Biochem. Pharm.* **1988**, 37, 2987. Cushman, M.; Nagarathnam, D.; Geahlen, R. L. *J. Nat. Prod.* **1991**, 54, 1345. Cushman, M.; Zhu, H.; Geahlen, R. L.; Kraker, A. J. *J. Med. Chem.* **1994**, 37, 3353. Miller, D.; Wang, S.; Reid, J.; Xie, W.; Gauvin, B.; Kelley, M.; Sarup, J.; Sawutz, D. G.; Miski, M.; Dolle, R. E.; Faltynek, C. R. *Drug Development Res.* **1995**, 34, 344.
5. Burke, T. R.; Ford, H.; Oshero, N.; Levitzki, A.; Stefanova, I.; Horak, I. D.; Marquez, V. E. *Bioorg. Med. Chem. Lett.* **1992**, 1771. Smyth, M. S.; Stefanova, I.; Hartmann, F.; Horak, I. D.; Oshero, N.; Levitzki, A.; Burke, J. T. R. *J. Med. Chem.* **1993**, 3010. Smyth, M. S.; Stefanova, I.; Horak, I. D.; Burke, J. T. R. *J. Med. Chem.* **1993**, 3015. Showalter, H. D. H. *Chemtracts* **1993**, 258.
6. Faltynek, C. R.; Schroeder, J.; Mauvais, P.; Miller, D.; Wang, S.; Murphy, D.; Lehr, R.; Kelley, M.; Maycock, A.; Michne, W.; Miski, M.; Thunberg, A. L. *Biochemistry* **1995**, 34, 12404.
7. Hanke, J. H.; Gardner, J. P.; Dow, R. L.; Changelian, P. S.; Brissette, W. H.; Weringer, E. J.; Pollock, B. A.; Connelly, P. A., *J. Biol. Chem.* **1996**, 271, 695.
8. Barker, A.; Davies, D. H., EP Appl. 520722, 1992. Barker, A. J., EP Appl. 566226, 1993. Barker, A. J.; Brown, D. S., EP Appl. 602851, 1994. Barker, A. J., EP Appl. 635498, 1995. The EGF-R activity of 4 has been reported previously: Levitzki A, Gazit A. *Science* **1995**, 267, 1782.
9. (a) Fry, D. W.; Kraker, A. J.; McMichael, A.; Ambrosio, L. A.; Nelson, J. M.; Leopold, W. R.; Connors, R. W.; Bridges, A. J. *Science* **1994**, 265, 1093. (b) Bridges, A. J. *Exp. Opin. Ther. Patents* **1995**, 5, 1245 and references therein. (c) Bridges, A. J.; Cody, D. R.; Zhou, H.; McMichael, A.; Fry, D. W. *Bioorg. Med. Chem.* **1995**, 3, 1651.
10. Traxler, P. M.; Furet, P.; Mett, H.; Buchdunger, E.; Meyer, T.; Lydon, N. *J. Med. Chem.* **1996**, 39, 2285.
11. (a) Spada, A. P.; Persons, P. E.; Levitzki, A.; Gilon, C.; Gazit, A. WO PCT 91/16501, publ. Oct. 31, 1991. (b) Spada, A. P.; Maguire, M. P.; Persons, P.; Myers, M. R., WO PCT 92/20642, publ. Nov. 26, 1992. (c) Maguire, M.; Sheets, K. R.; McVety, K.; Spada, A. P.; Zilberstein, A. *J. Med. Chem.* **1994**, 37, 2129.
12. Niementowski, S. *J. Prakt. Chem.* **1895**, 51, 564.
13. Kreutzberger, A.; Uzbek, M. U. *Arch. Pharm.* **1972**, 305, 171.
14. For the p56^{lck} tyrosine kinase assay, compounds were screened initially at concentrations of 10 and 100 μ M in reactions containing p56^{lck} kinase immunoprecipitated from Jurkat cell lysates, 5 μ M cdc2 [a p34cdc2-derived synthetic peptide (N6-20)] substrate, 5 mM MnCl₂, 5 μ M ATP and 1 μ Ci [³²P] γ -ATP in 20 mM HEPES buffer (pH 7.5) for 5 min at 30 °C. Samples were analyzed by 5-15% SDS-PAGE and autoradiographs quantitated by densitometry. For the EGF-R assay see ref. 11b. In all cases, IC₅₀'s were from a minimum of two separate experiments.
15. See following communication in this journal.
16. Experiments performed by Dr. E. Rabin, RPR.